

# Understanding oscillations in gas fermentation

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## HIGHLIGHTS:

- Metabolism in a biological system is ultimately determined by the organisms' ability to self-organise metabolism for maintaining cellular homeostasis.
- Quantification of the limits of metabolic robustness is essential for understanding cellular behaviour
- This work can contribute towards advancing the understanding of an industrially relevant cell factory used in gas fermentation for sustainable production of fuels and chemicals from waste feedstocks.

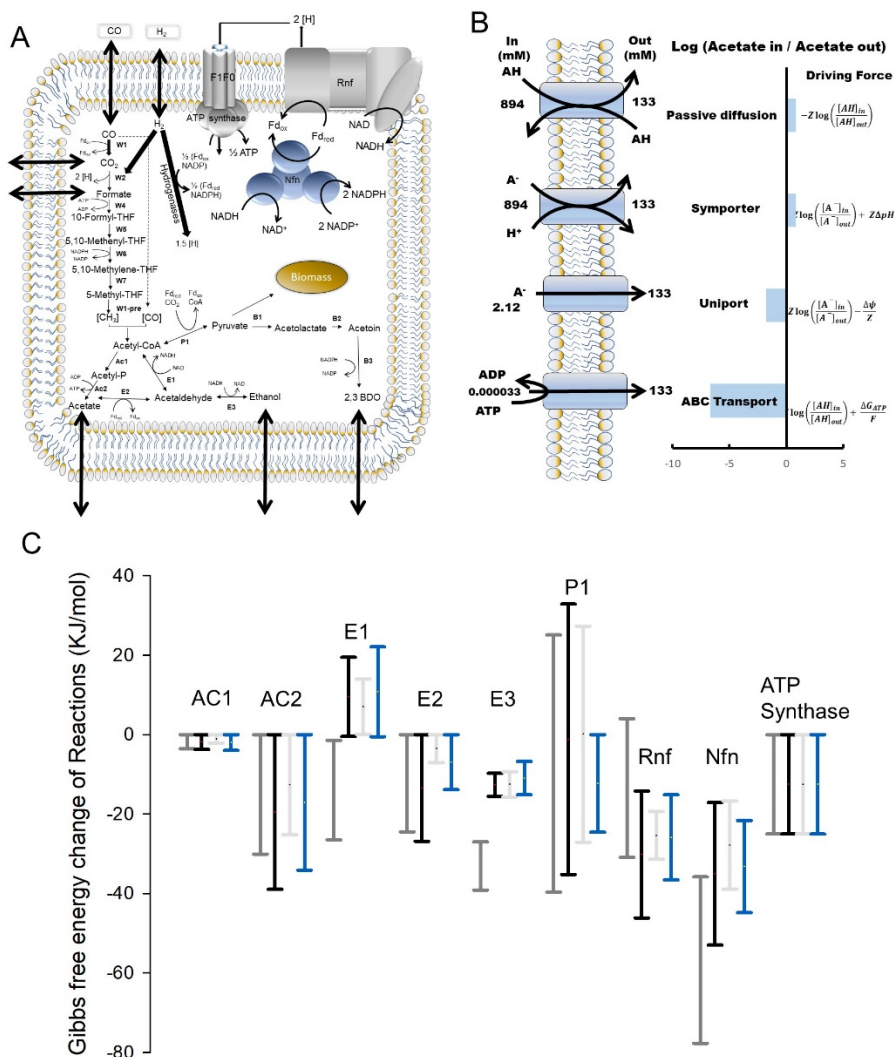
**BACKGROUND:** Gas fermentation can play a crucial role in the circular economy. However, oscillations have been observed in gas fermentation. Oscillatory behaviour provides a unique opportunity for quantitating the robustness of metabolism, as cells respond to changes by inherently compromising metabolic efficiency.

**RESULTS & DISCUSSION:** Here, we quantify the limits of metabolic robustness in self-oscillating autotrophic continuous cultures of the gas-fermenting acetogen *Clostridium autoethanogenum*. On-line gas analysis and high-resolution temporal metabolomics showed oscillations in gas uptake rates and extracellular by-products synchronised with biomass levels. We found that the intrinsic nature of gas fermentation allows for an

39 initial growth phase on CO, followed by growth on CO and H<sub>2</sub>, after which a  
 40 downcycle is observed in synchrony with a loss in H<sub>2</sub> uptake. Intriguingly,  
 41 oscillations are not linked to translational control as no differences were  
 42 observed in protein expression during oscillations. Intracellular  
 43 metabolomics analysis revealed decreasing levels of redox ratios in  
 44 synchrony with the cycles. We used a thermodynamic metabolic flux  
 45 analysis (tmFA) model to investigate if regulation in acetogens is controlled  
 46 at the thermodynamic level. By incorporating endo and exo-metabolomics  
 47 data into the model we could show that the thermodynamic driving force of  
 48 critical reactions collapsed as H<sub>2</sub> uptake is lost.

49 **CONCLUSION:** The oscillations are coordinated with redox. The data  
 50 indicate that metabolic oscillations in gas fermentation acetogens are  
 51 controlled at the thermodynamic level. The work presented will show that  
 52 thermodynamic control of metabolism, potentially contributing to metabolic  
 53 efficiency in gas fermentation.

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56 Figure 1. Thermodynamic metabolic flux analysis (tMFA) of *Clostridium*  
57 *autoethanogenum*. (A) Schematic representation of our tMFA model of *C.*  
58 *autoethanogenum*. Arrows across the cell membrane denote product  
59 transport. (B) Modelling four mechanisms for acetate transport which were  
60 then used in our tMFA model. See text and Excel datasheet. 2,3-BDO, 2,3-  
61 butanediol; THF, tetrahydrofolate; Fd, ferredoxin; A<sup>-</sup>, acetate; AH, acetic  
62 acid; H<sup>+</sup>, proton. (C) The plot shows the maximum allowable range of Gibbs  
63 free energy for respective reactions at given conditions calculated using  
64 thermodynamic variability analysis (TVA); Grey and black show the three  
65 phases from this work and blue shows data from the steady state. See text  
66 for details; Each bar denotes mean and error bars standard deviation of  
67 that allowable range. \* Positive reactions are thermodynamically not  
68 feasible at given conditions (Have positive Gibbs free energy change)

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## 70 REFERENCES

- 71 1. Mahamkali, et. al. Redox controls metabolic robustness in the gas-  
72 fermenting acetogen *Clostridium autoethanogenum*.  
73 PNAS 2020, 117 (23) 13168-13175; DOI: 10.1073/pnas.1919531117