# Production of carboxylates (C-A)<sup>1</sup> by young sugar-beet plants grown in nutrient solution

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#### Summary

An experiment was carried out with sugar-beet grown in a nutrient solution to investigate whether decarboxylation takes place in the roots of young sugar-beet plants. Decarboxylation, balanced by uptake of anions in excess of cations from the nutrient solution, were not detected. Only a small amount of carboxylates reached the root system. The reduction of nitrate was almost equal to the carboxylate production. This reduction of nitrate per gram of leaf per day decreased rapidly with time. The supposition is made that the high oxalate content in the leaves might account for the decrease in nitrate reduction.

# Introduction

The uptake of nitrate nitrogen by plant roots and its reduction within the plant results in production of an equivalent amount of carboxylates (Dijkshoorn, 1968). In experiments with perennial rye-grass (*Lolium perenne* L.), Dijkshoorn recovered only 1 eq of carboxylates of the 2.5 eq originally produced by the reduction of 2.5 eq of nitrate per kg of dry matter. The missing amount of 1.5 eq was decarboxylated (probably in the root), the bicarbonate released being balanced by uptake of anions in excess of cations from the nutrient solution.

Houba's data (unpublished) on sugar-beet (*Beta vulgaris* L.), sufficiently supplied with nitrate, show that the nitrate equivalent of organic nitrogen equals the amount of carboxylate equivalents. His data refer to sugar-beet plants grown in a field experiment and harvested when the formation of beets had already taken place.

An experiment was set up to investigate whether young sugar beet plants will show the process of decarboxylation supposed to take place in perennial rye-grass. This experiment was carried out with nutrient solution, during the period in which practically no beets are formed.

# Experimental

Diploid sugar-beet seeds (No P2167) were sown in a thin layer of gravel on sieves that were placed on top of one-litre polyethylene pots. After germination the roots grew through the gravel on the sieve and entered a well-aerated nutrient solution.

<sup>&</sup>lt;sup>1</sup> The term (C-A) represents the amount of inorganic cations minus that of inorganic anions both expressed in meq per kg of dry matter.

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Component	Concentration (meq/l)
Na	2
К	4
Ca	2.5
Mg	4
H <sub>2</sub> PO <sub>4</sub>	1
NO <sub>3</sub>	6
Cl	3
SO4	2.5
Trace elements (except Fe)	according to Hoagland
Fe-EDTA	35 mg/1

Table 1	Chemical	composition	of	the	nutrient	solution	
rable r	Chemicai	composition	U	ine	питет	solution	

The composition of the nutrient solution is given in Table 1. To cope with expected shortages the nutrient solutions were renewed at regular intervals. One plant was grown in each pot. The experiment was carried out in a climate-controlled growth cabinet. The daily light period was 14 hours, the temperature was  $25 \,^{\circ}$ C during the light period and  $17 \,^{\circ}$ C during the dark period, the relative humidity was 80 to  $90 \,^{0}/_{0}$ . Plants were harvested after 30, 36, 39, 43, 46 and 51 days counting from the day of germination, and divided into three sections: leaf blades (including the midribs), petioles and beet + roots. Of each section fresh and dry weights were recorded and the material was finely ground and analysed for Na, K, Ca, Mg, H<sub>2</sub>PO<sub>4</sub>, NO<sub>3</sub>, Cl, SO<sub>4</sub>, total N, organic acids (i.e. oxalic acid, citric acid, fumaric acid, succinic acid and malonic acid).

Subsamples were analysed after digestion in concentrated sulphuric acid and hydrogen peroxide in the presence of salicylic acid. In this digest, the Na, K and Ca contents were determined flamephotometrically, the  $H_2PO_4$  content colorimetrically, the Mg content by atomic absorption and the nitrogen content (total N) by the micro-Kjehl-dahl procedure. Other subsamples were extracted with water (0.5 g of plant material and 50 ml deionised water: shaking time 2 hours). In this extract the nitrate-content was determined with an Orion nitrate electrode, the Cl content was determined coulo-metrically with a chloro counter and the SO<sub>4</sub> content colorimetrically. The total content of carboxylates (C–A) was calculated by subtracting the sum of inorganic anions from the sum of inorganic cations (de Wit et al., 1963). The carboxylates were converted into organic acids by decationization with a H<sup>+</sup> sulphonic acid resin and these were resolved by partition chromotography and quantitatively determined with an automatic titrator. All values were calculated on the basis of oven-dried material (70° C).

#### **Results and discussion**

#### Dry-matter production

During the experiment the dry weights showed a lineair increase in dry matter with time (see Fig. 1A). From the fourth harvest onwards the weight of roots (including the beet) expressed in percentage of the total dry weight, increased significantly (Table 2). This indicates the beginning of the stage of beet formation.

### The balance of uptake and utilisation of inorganic ions

The most important components which may characterize the balance of uptake and

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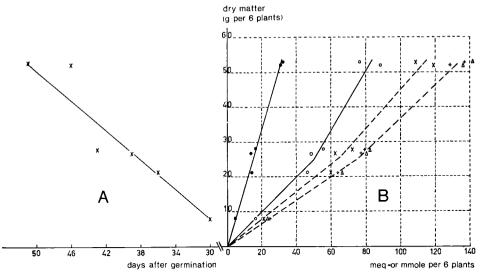


Fig. 1 A. Production of dry matter; B. organic N, (C-A) and carboxylates; • mobile acids; • oxalic acid;  $\times$  carboxylates; + organic N;  $\triangle$  (C-A)

utilisation of ions, are given in Table 4 and Fig. 1B. The table shows that the figures for (C-A), and for organic nitrogen are about equal (difference  $\pm 3 \circ/_0$ ). Generally the amount of organic nitrogen is somewhat lower than the amount of (C-A) (organic N expressed as its ion equivalent of nitrate). Considering the amount of organic sulphur the (C-A) is somewhat lower than the sum of organic N and organic S. (According to Dijkshoorn and van Wijk (1967) organic S is about  $6 \circ/_0$  of organic N on ion equivalent basis.)

Since the amounts of (C-A) and organic N are about equal, there is no release of carboxylates resulting from  $NO_3^-$  and  $SO_4^{2^-}$  reduction in the leaves in sugar-beet plants and consequently no excess uptake of inorganic anions over inorganic cations. Of the total amount of (C-A) formed by reduction of nitrate and sulphate, only  $6-8 \ 0/0$  is present in the beet-root system. When only the roots represent the uptake system, a part only of the  $6-8 \ 0/0$  mentioned above reaches this uptake system. So the process Dijkshoorn et al. (1968) supposed to take place in perennial rye-grass does not take place in sugar-beet plants, or could not be demonstrated in sugar-beet plants because of the very small amount of carboxylates that reaches the uptake system.

Table 2 Dry-weight production (g/6 plants) of sugar-beet plants and the distribution of the produced dry weight over leaf blade, petiole and roots (beet included)

Days after germination	Leaf blade	Petiole	Root and beet	Total	Roots (% of total dry weight
30	5.32	1.4	1.24	7.96	16
36	13.6	4.2	3.6	21.4	17
39	16.6	5.4	4.8	26.8	18
43	17.6	5.6	4.8	28.0	17
46	29.0	10.6	12.6	52.2	25
51	28.8	10.65	13.65	53.1	26

410       177       68         530       197       72         480       168       64         570       270       76         657       169       66         275       169       66         1900       883       125         1900       883       125         1820       801       86         1760       803       125         1820       1063       69         1760       81       125         1820       1063       69         720       346       35         810       354       35         720       346       35         720       346       35         730       301       51         730       301       51         370       374       37         730       301       51	197 197 146 162 152 154 154 154 153 133 133 133 140 154 154 154 154 154 154 154 153 133 133 133 140	4801 4801 4957 4775 4775 4775 4822 4822 4822 4802 4177 4177 4177 3318 3318 3318 3318 3318 3318 3318 3	
53		2167         227         570           1777         242         376	1520         344         2167         227         570           1130         334         1777         242         376

Days after	(C-A)	Organic N	Carboxylates	
germination	(meq)	(mmole)	(meq)	(% of (C-A))
30	24.8	24.3	21.0	84.7
36	64.0	64.1	60.1	93.9
39	78.8	76.4	62.3	79.1
43	80.0	78.4	71.9	89.9
46	132.8	128.7	119.4	89.9
51	141.9	136.9	108.7	76.6

Table 4 Organic N, (C-A) and carboxylates in the combined plant parts

A reason for this relatively low amount of carboxylates reaching the root system is perhaps the very low solubility of a large portion of these carboxylates. Illustrative for the situation in sugar-beet plants are the high contents of Ca and Mg (Table 3) together with high contents of oxalates (the solubility of these oxalates is low).

The change in slope of the lines in Fig. 1B at the fourth harvest corresponds with the moment at which the plants start forming beets (Table 2). In the first period of growth when no beets are formed, on the average 3000 mmole of organic N and 2000 meq of oxalate are produced per kg of dry matter. During the second growth period when formation of beets takes place, these values are 2100 mmole and 1200 meq, respectively. The differences in organic nitrogen and oxalate content between the two periods amount to 900 mmole and 800 meq per kg of dry matter, respectively. The rate of production of 'mobile' carboxylates — mostly citrate and malate — does not change and is maintained at a value of about 600 meq per kg of dry matter.

# Nitrate reduction

From the data on leaf-blade weights (Table 2), one can calculate an average production of about 1.1 g of leaf blade per 6 plants per day. In the same way one can calculate an average  $NO_3$  reduction of about 6.0 meq  $NO_3$  per 6 plants per day (Table 4: data on organic N). On the 30th day after germination, there are 5.3 g leaf blades per 6 plants. With these data the calculations given in Table 5 can be made. Every value recorded in Fig. 2 is based on a calculation as given above. From this figure it is clear that nitrate reduction decreases rapidly with time. This decreasing nitrate reduction in the presence of sufficient amounts of available nitrate in the tissue (Table 3) results in a decreasing production of oxalate (Fig. 1B). Whether a lower nitrate reduction causes a lower oxalate production or whether an accumulation of oxalate causes a lower nitrate reduction was not investigated. It seems, however, quite

Days after germination	Leaf blade (g) per 6 plants	Average weight of leaf blade (g)	NO3 <sup>-</sup> reduction per gram leaf blade per day (meq)
30	5.3	5.85	$\frac{6.0}{5.85}$ = 1.03
31	5.3 + 1.1	6.95	6.0
32	6.4 + 1.1	0.75	6.95

 Table 5
 Calculation of average weight and NO3<sup>-</sup> reduction of leaf blades

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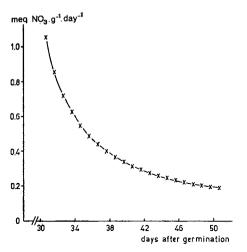


Fig. 2 Reduction of nitrate (meq) per leaf blade per day (calculated from produced organic N and produced dry matter of leaves)

likely that there is a maximum to the quantity of oxalate a leaf can contain. From the calculated quantity of carboxylates (C–A), 85  $^{0}$ /<sub>0</sub> is recovered in the organicacid analyses (Table 4). The missing 15  $^{0}$ /<sub>0</sub> cannot be explained with the water-soluble carboxylates. Van Tuil (1965) assumes the presence of other carboxylates which are difficult to extract.

# Conclusions

In contrast with rye-grass, an excess uptake of inorganic anions over inorganic cations accompanied by a decarboxylation of carboxylates, could not be demonstrated in young sugar-beet plants. This could be the result of a relative low amount of carboxylates reaching the roots.

In sugar-beet plants the (C-A) content is not constant during the growth period studied in this experiment.

The nitrate reduction per gram leaf per day decreases rapidly with time.

The decrease in nitrate reduction is accompanied by decrease in oxalate production.

# Acknowledgment

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