# Denitrification in nitric-acid-treated cattle slurry during storage

O. OENEMA & G.L. VELTHOF

NMI, Department of Soil Science and Plant Nutrition, Wageningen Agricultural University, P.O. Box 8005, NL 6700 EC Wageningen, Nether-lands

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### Abstract

Treatment of cattle slurry with HNO3 to lower the pH is a measure to reduce NH3 volatilization from this slurry during storage and after surface application. Moreover, the addition of HNO3-N increases the fertilizer value of the slurry. Incubation studies were carried out to examine possible NO<sub>3</sub> losses and N<sub>2</sub>O emission from HNO<sub>3</sub> treated slurry during storage. Batches of cattle slurry were treated with various amounts of HNO3 to obtain a pH range of 6.0 to 3.0. The slurries were stirred once or twice a week and stored for 6 months at 15 °C. Changes in pH, Eh, concentrations of NO<sub>3</sub> and NH<sub>4</sub>, and emissions of N<sub>2</sub>O, CO<sub>2</sub> and CH<sub>4</sub> were monitored. The loss of NO<sub>3</sub> and the emission of N<sub>2</sub>O were related to slurry pH, being lowest at low pH, Cumulative loss of NO3 ranged from about 40 mmol kg-1 for slurries of target pH ≤ 5.0 to about 400 mmol kg-1 for slurries of target pH 6.0. Homogenization of the slurries via stirring and addition of H2O2 decreased the NO3 loss and H+ consumption. The strong pH-dependance of the NO<sub>3</sub> loss, the production of N<sub>2</sub>O and the stoichiometry of H<sup>+</sup> consumption and NO3 loss indicated that the loss of NO3 was mainly due to microbiological denitrification. Similar N2O production rates in the presence and absence of C2H2 indicated that nitrification was not an important source of N2O. It is concluded that lowering the pH to values ≤ 4.5 as well as regular and complete homogenization of the slurry via stirring are important for the success of the on-farm treatment of slurry with HNO3.

Keywords: nitrous oxide, ammonia volatilization, proton consumption, redox potential, pH

#### Introduction

In the past few years a number of measures has been introduced to reduce ammonia (NH<sub>3</sub>) emission from cattle slurries on Dutch dairy farms. Such measures include immediate storage of slurry in proper covered basins, application through injection into the soil and feeding the cattle an adjusted ration (e.g. Den Boer et al., 1990). In practice, a number of measures often have to be combined, to ensure that all entrapped NH<sub>3</sub> can be used for plant growth and to prevent that entrapped N is lost via other pathways in the N cycle.

An alternative measure that may integrate the effect of a number of measures is

the treatment of cattle slurry with acid to pH 4.0-4.5, immediately after its deposition by the animal in the livestock building. Due to the shift in the NH<sub>3</sub>:NH<sub>4</sub><sup>+</sup> equilibrium, the emission of NH<sub>3</sub> from acid treated slurry is reduced during storage, and during and after surface application. Field experiments have shown indeed that NH<sub>3</sub> losses from such slurries are much lower than from untreated slurry after surface application (e.g. Stevens et al., 1989). If treated with nitric acid (HNO<sub>3</sub>), the fertilizer value of the slurry also increases, strongly reducing the need for the additional use of mineral N fertilizers.

Unfortunately, a mixture of cattle slurry and HNO<sub>3</sub> is not very stable. Cattle slurries are highly anoxic and under such conditions the added NO<sub>3</sub> may be denitrified to nitrous oxide (N<sub>2</sub>O) and N<sub>2</sub>, even though the low pH will strongly limit the rate of the microbial denitrification (Knowles, 1982). The trace gas N<sub>2</sub>O possibly contribute to global warming (Wang et al., 1976) and the catalytic destruction of stratospheric ozone (Crutzen, 1976). Denitrification also increases the pH of the slurry, so that NH<sub>4</sub> may be deprotonated into NH<sub>3</sub> and then still be emitted into the atmosphere. Control of denitrification seems therefore crucial for a successful on-farm implementation of the treatment of cattle slurries with HNO<sub>3</sub>.

Thus far, pilot farms have faced variable NO<sub>3</sub> losses from HNO<sub>3</sub> treated slurries of pH 4.0-4.5 during storage. Variations in slurry composition, storage time and temperature, and insufficient mixing leading to variations in slurry pH have been put forward as major factors accounting for the variable NO<sub>3</sub> loss. This prompted us to study the effects of pH, slurry mixing and storage time on denitrification and trace gas emissions from HNO<sub>3</sub> treated slurries during storage and after surface application. This paper mainly deals with the NO<sub>3</sub> losses from HNO<sub>3</sub> treated slurries during storage under semicontrolled conditions. Emissions of N<sub>2</sub>O after surface application of treated and untreated slurries are summarized by Velthof & Oenema (1993).

We examined the effect of pH values ranging from 3.0 to 6.0 on NO<sub>3</sub> loss from slurry during storage. This range is much wider than the range of pH 4.0 to 4.5 which has been proposed for the on-farm treatment and storage of slurry.

### Materials and methods

# Titration curves of cattle slurries

Titration curves were established by adding 0.25 to 0.5 ml of 10.3 M HNO<sub>3</sub> to 200 g slurry under continuous stirring. Once the pH stabilized, usually after 5 to 10 min, another 0.25 to 0.5 ml HNO<sub>3</sub> was added until the pH was 3.0. Reverse titration was carried out by intermittently adding 0.25 to 0.5 ml of 4.0 M NaOH to 200 g slurry under continuous stirring.

## Incubation of cattle slurries

Effects of pH, mixing and storage time on NO<sub>3</sub> losses and N<sub>2</sub>O emission from HNO<sub>3</sub> treated cattle slurry were studied in a batch experiment with an incomplete randomized block design.

Fresh cattle slurry of less than one week old was obtained from a local dairy farm. Homogenized samples of 12 kg each in 38 PVC containers (26 l) were acidified to pH 6.0, 5.0, 4.5, 4.0, 3.5 and 3.0 by slow addition of 10.3 M HNO<sub>3</sub> under moderate stirring. A control treatment with no addition of HNO<sub>3</sub> was included. At all 7 pH levels, there were three stirring treatments in duplicate, except for pH levels 3.5 and 3.0, which were combined with two stirring treatments only. Slurries of stirring treatment A were gently stirred with a rudder blade at 90 revolutions per min, for 2 min once a week. This allowed sampling from homogeneous slurries. Slurries of treatment B were heavily stirred at 240 revolutions per min, for 5 min twice a week. Slurries of treatment C received as oxidant 7.5 mmol H2O2 kg-1 twice a week and were stirred as B. Oxidation of some of the easily metabolizable organic compounds in the slurry by H2O2 was assumed to diminish the amount of substrate for denitrifiers and hence the loss of NO3. Combinations of treatment C and pH levels 3.5 and 3,0 were excluded. The purpose-designed rudder blades allowed a rapid and complete mixing of all slurry. Containers with slurry were incubated in a climate-conditioned room at 15±0.5 °C for 6 months from January to August 1991.

Denitrification rates in the slurries were deduced from the loss of NO<sub>3</sub> and consumption of protons (H<sup>+</sup>) in the course of time. Additional measurements of denitrification rates were carried out, using the acetylene inhibition technique (Ryden & Rolston, 1983). Acetylene (C<sub>2</sub>H<sub>2</sub>) is an inhibitor of nitrification (Bremner & Blackmer, 1979) and of the reduction of N<sub>2</sub>O to N<sub>2</sub>, the last step in the denitrification process (Yoshinari & Knowles, 1976). Portions of 10 ml slurry were incubated under ambient air and under ambient air with 10 % acetylene in glass vials with rubber septa. Gas samples of 0.5 ml were taken every 30 min from the head space, and were analyzed for N<sub>2</sub>O.

The pH of the incubated slurries was monitored once or twice a week, and the Eh once every two weeks. HNO<sub>3</sub> was added to the slurries if the actual pH exceeded the target pH by more than 0.15 units. Concentrations of dissolved and exchangeable NH<sub>4</sub> and NO<sub>3</sub> in the slurries were measured in 20-ml subsamples, after homogenization by stirring and correction of slurry pH if necessary. The subsamples were transferred to 50-ml polypropylene tubes containing 25 ml 1.0 M KCl. After shaking for 15 min, the slurries were centrifuged at 4000 revolutions per min for 15 min. The supernatant was diluted 100 to 250 times and analyzed by means of Technicon autoanalyzer. N<sub>2</sub>O concentrations in the slurries were determined in 5-ml subsamples transferred into 70-ml glass vials with rubber caps. Vials were strongly agitated for 5 min and the concentration increase in the headspace was analyzed via gas chromatography.

Fluxes of N<sub>2</sub>O and CO<sub>2</sub> from incubated slurries were calculated from the changes in the N<sub>2</sub>O and CO<sub>2</sub> concentrations in the headspace after the containers were sealed, usually for 0.5 to 2 h. Gas samples were taken with 12-ml polypropylene syringes through rubber septa in the caps at regular time intervals. Samples were analyzed by gas chromatography. During the last month we also used a photo acoustic infra-red Multi-gas Monitor 1302 from Brüel & Kjær, directly attached to the containers (cf. Velthof & Oenema, 1993).

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# Analytical procedures

Redox potential (Eh) and pH were measured electrochemically.  $NO_3^-$  and  $NH_4^+$  were analyzed by standard auto-analyzer methods (Technicon; US no 824-87T and US no 795-86T, respectively). The  $NO_3^-$  analyses were reasonably accurate; the linear relationship between the amount of  $NO_3^-$  added (x) via  $HNO_3$  and the amount of  $NO_3^-$  analyzed (y) in 42 samples was: y = 0.03 + 0.99x with  $r^2 = 0.995$ . Total C, N, P and K in the slurry were analyzed using standard methods (Vierveijzer et al., 1979).  $N_2O$  and  $CO_2^-$  in 0.5 ml gas samples were analyzed via a PU 4400 gas chromatograph using a 27.5 m Porapak Q wide bore column and an electron capture detector (ECD).

#### Results

# Changes in overall slurry composition

The fresh slurry, a mixture of urine and dung, had a relatively high pH and high total N and K contents compared to average values for cattle slurry (Table 1). Organic C content of the solids was about 42% and the molar ratio of total C:N:P was about 101:13:1. During storage, the weight of the slurries decreased by about 14% due to evaporation. To facilitate comparison, contents of incubated slurries were corrected for weight loss by evaporation. Total solids, organic C, total N and NH<sup>+</sup><sub>4</sub> contents of untreated slurry decreased during storage due to decompostion of organic matter and subsequent volatilization of CO<sub>2</sub>, NH<sub>3</sub> and possible other volatile compounds like fatty acids and H<sub>2</sub>S. The pH of the untreated slurries gradually decreased from 8.4 to about 7.5 within 30 days, possibly as the result of NH<sub>3</sub> volatilization and precipitation of carbonate and struvite (Bril & Salomons, 1990), and remained at that level for the remaining 145 days.

Addition of HNO<sub>3</sub> slightly increased the total volume and the content of total solids of the slurry. The treatment with HNO<sub>3</sub> had a significant effect on the C con-

Table 1. Average composition of the cattle slurry at the start of the experiment and after incubation for six months at various pH levels (g kg<sup>-1</sup>). The composition of incubated slurries was corrected for weight loss due to evaporation.

|                    | Fresh<br>slurry | Slurries after six month incubation |                           |          |     |  |  |
|--------------------|-----------------|-------------------------------------|---------------------------|----------|-----|--|--|
|                    | siuity .        | Untreated                           | HNO <sub>3</sub> -treated |          |     |  |  |
| pH                 | 8.4             | 7.5                                 | 6.0                       | 4.5      | 3.0 |  |  |
| pH<br>Total solids | 8.4<br>113      | 88                                  | 118                       | 132      | 133 |  |  |
| Organic C          | ' 47            | 38                                  | 38                        | 42       | 45  |  |  |
| NH <sub>4</sub> -N | 4.3             | 3.0                                 | 3.4                       | 4.1      | 4.1 |  |  |
| NO3-N              | 0.0             | 0.0                                 | 4.9                       | 6.5      | 7.0 |  |  |
| Total N            | , 7.3           |                                     | _                         | _        | _   |  |  |
| Total P            | 1.2             | _                                   | <u> </u>                  |          |     |  |  |
| Total K            | 7.1             | _                                   | <del>-</del>              | <u> </u> | _   |  |  |

<sup>\*</sup> Not determined.

tent of the slurry after incubation for 6 months. Stirring did not have a distinct effect, so that averaged values of treatments A, B and C are given in Table 1 for selected pH-levels only. After storage for 6 months, the organic C and NH<sup>+</sup><sub>4</sub>-N contents of slurries of pH ≤5.0 were only slightly decreased relative to the fresh slurry, suggesting that rates of organic matter decompostion and NH<sub>3</sub> volatilization were low. However, organic C and NH<sup>+</sup><sub>4</sub>-N contents were significantly decreased in the HNO<sub>3</sub> treated slurry of target pH 6.0.

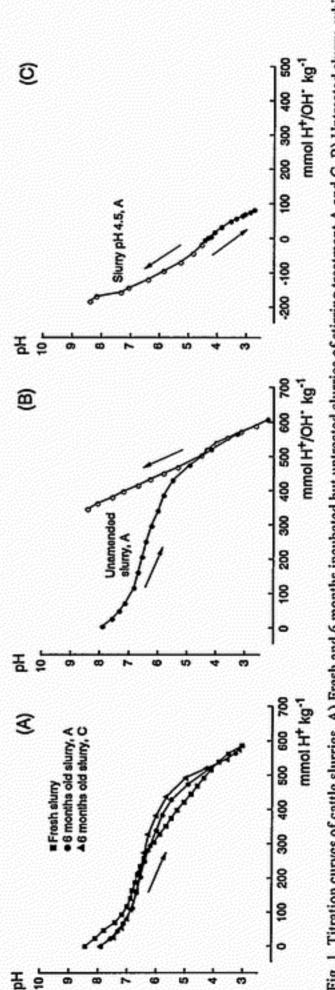
# Effects of HNO3 addition on slurry pH and Eh

The relationship between added amounts of acid (H<sup>+</sup>) and base (OH<sup>-</sup>) and the pH of the slurry is shown in Fig. 1. The slurry had a high acid neutralizing capacity (ANC). The ANC of the slurry was highest within the range of pH 7.0 to 6.0. Ageing of untreated slurries slightly decreased the ANC at pH ≥6.0, possibly as a result of NH<sub>3</sub> volatilization, and increased the ANC in the range of pH 6.0-5.0. Total ANC was little affected during storage. The base neutralizing capacity (BNC) of slurry that was acidified to pH 2.5 was less than half of the ANC of untreated slurry (Fig. 1B). This suggests that a large fraction of the ANC was associated with (bi)carbonate and anions of fatty acids (Husted et al., 1991; Japenga & Harmsen, 1990), which volatilized as CO<sub>2</sub> and volatile fatty acids after H<sup>+</sup> addition. The BNC was about 42 mmol kg<sup>-1</sup> per unit pH change and was essentially constant between pH 2.5 and 7. Once the ANC associated with volatile compounds had escaped, the titration and reverse titration curves were similar (Fig. 1C), provided that care was taken not to trap CO<sub>2</sub> in the slurry at high pH.

Addition of HNO<sub>3</sub> increased the Eh of the slurry. Untreated slurry was highly anoxic and had a mean Eh of about -400 mV (Fig. 2). The Eh of HNO<sub>3</sub> treated slurries was ranging from 0 to 300 mV. More intensive stirring (treatment B) and  $H_2O_2$  addition (treatment C) had no distinct effect on the Eh of untreated slurry and HNO<sub>3</sub> treated slurry of target pH 6, but significantly increased the Eh of HNO<sub>3</sub> treated slurries of pH  $\leq$  5.0. Effects of stirring became most clear after about 40 days (Fig. 2); after the initial 'adaption phase' the Eh remained relatively constant. Even though Eh was increased by stirring and the addition of HNO<sub>3</sub> and  $H_2O_2$ , it did not surpass 400 mV, a redox potential which others have found as an upper limit for denitrification (e.g. Letey et al., 1982).

# Rates of H<sup>+</sup> and NO<sub>3</sub> consumption

A typical course of the actual slurry pH during storage is shown in Fig. 3. The pH of HNO<sub>3</sub> treated slurries of target pH < 5.0 remained relatively constant for most of the storage time. Addition of H<sub>2</sub>O<sub>2</sub> (treatment C) slightly decreased the pH, suggesting the release of H<sup>+</sup> through the oxidation of reduced inorganic compounds, like H<sub>2</sub>S and NH<sub>4</sub>. A notable increase in pH was observed after day 125, irrespective of stirring. This increase was probably associated with the gradual increase of the incubation temperature from 15±0.5 °C until day 125 to about 22 °C during the last month. The reason for this increase was that the climate conditioner could not cope with the high out-



was incubated for 6 months, titrated with HNO3 to pH 2.5 and reverse with NaOH. C) Titration and reverse titration of HNO3 treated slurry of pH 4.5, Fig. 1. Titration curves of cattle slurries. A) Fresh and 6 months incubated but untreated slurries of stirring treatment A and C. B) Untreated slurry, which which was incubated for 6 months.

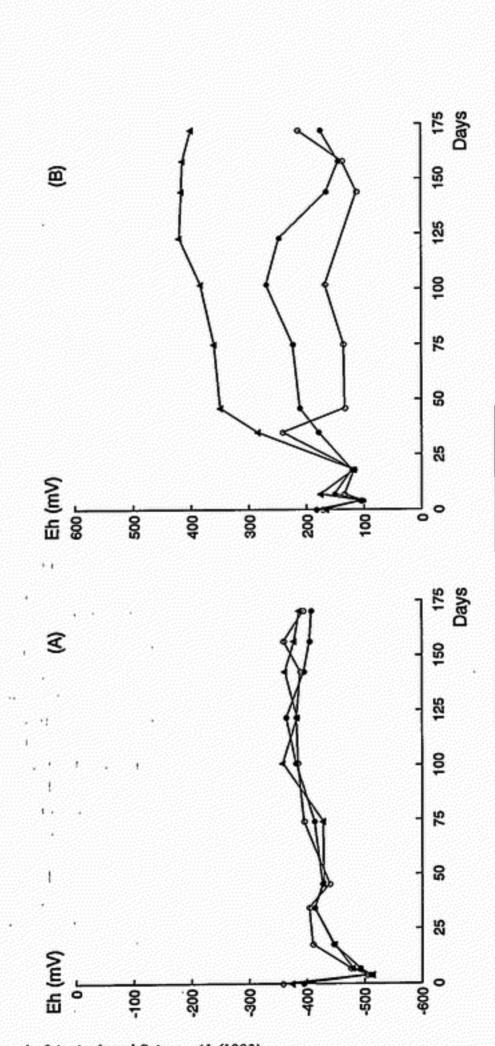


Fig. 2. Changes in redox potential (Eh) of slurry during incubation for 6 months. A) Untreated slurry. B) HNO<sub>3</sub> treated slurry of pH 4.5. Note the scale differences of the y-axis.

◆ Treatment C

Treatment A • Treatment B

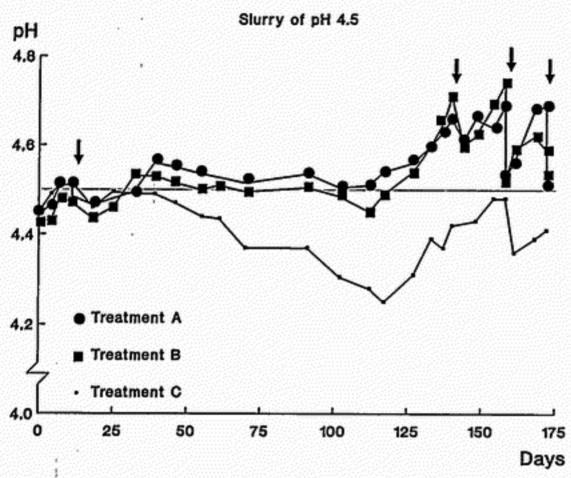


Fig. 3. Changes in pH of HNO<sub>3</sub> treated slurry of target pH 4.5, as a function of stirring treatments A, B and C, and incubation time. Arrows indicate pH correction by HNO<sub>3</sub> addition.

side temperature during the hot summer months of 1991. As a result, more than 75% of the total H<sup>+</sup> consumption in slurries with target pH < 5.0 occurred after day 125.

Slurries of target pH 6.0 had a high and nearly constant H<sup>+</sup> consumption rate throughout the incubation period (Fig. 4). The mean H<sup>+</sup> consumption rate was more than 2 mmol kg<sup>-1</sup> d<sup>-1</sup> for slurries of stirring treatment A. More intensive stirring and addition of  $H_2O_2$  decreased the rate of H<sup>+</sup> consumption. The increase in H<sup>+</sup> consumption rate of slurries of stirring treatment A and B between day 112 and 138 was due to the fact that the pH was allowed to rise to about 8 between day 112 and 138. Such a rise in pH may have accelerated H<sup>+</sup> consumption rate. The temperature rise noted before may also have contributed. Table 2 summarizes the total H<sup>+</sup> consumption rate during incubation. Evidently, pH had a strong effect on H<sup>+</sup> consumption. H<sup>+</sup> consumption rate was doubled between pH ≤ 4.5 and 5.0 but increased tenfold between pH 5.0 and 6.0. Differences between stirring treatments A and B were relatively small, but addition of  $H_2O_2$  in treatment C significantly decreased H<sup>+</sup> consumption.

Concentrations of NO<sub>3</sub> were measured when the actual pH was equal to the target pH, after adjustment via HNO<sub>3</sub> addition if necessary. There were statistically

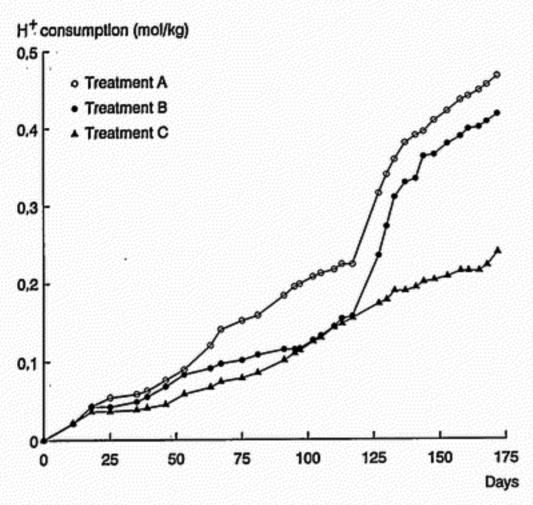


Fig. 4, Cumulative H<sup>+</sup> consumption by HNO<sub>3</sub> treated slurries of target pH 6.0 and stirring treatments A, B and C during incubation for 6 months.

significant effects of target pH and date on the NO3 and NH4 concentrations and there was a significant date × pH interaction (Table 2). Stirring had no significant effect on NO3 and NH4 concentrations. During incubation, NO3 concentrations increased in slurries of pH 6.0 and decreased in slurries of pH ≤ 4.0. Irregular variations were measured in slurries of pH 4.5 and 5.0. Changes in NO3 concentration were most apparent from the second month onwards. The overall mean NO3 loss was 8 mmol kg-1 between the first and last samplings. The significant effect of the date and the significant date x pH interaction for the NO3 data suggest that the H+: NO3 stoichiometry in the slurries slightly changed during incubation. However, as it follows from the Least Significant Difference (L.S.D.) in Table 2, the standard error of difference (s.e.d.) was rather large for the NO3 data. NH4 concentrations decreased in untreated slurries and in HNO3 treated slurries of pH 6.0 during incubation. This decrease was probably due to NH3 volatilization; it occurred predominantly during the first month, concomitant with the pH decrease in untreated slurry. The slightly higher NH4 losses in stirring treatment B than in treatment A suggest that the more intensive stirring in B has stimulated NH<sub>3</sub> volatilization. The total mean NH<sub>4</sub> loss was 17 mmol kg<sup>-1</sup>.

Flux of N2O, CO2 and CH4

Fluxes of CO2 and CH4 strongly decreased with a decrease in pH (Fig. 5), suggesting

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Table 2, Cumulative mean H<sup>+</sup> (HNO<sub>3</sub>) consumption and mean net changes in NO<sub>3</sub> and NH<sub>4</sub> concentrations (mmol kg<sup>-1</sup>) during incubation for 172 days, as a function of slurry pH and stirring treatments A, B, C. A summary of the analysis of variance of NO<sub>3</sub> and NH<sub>4</sub> concentrations is given in the second part of the table.

| pH            | H <sup>+</sup> co | nsumpti | on   | Change in [NO <sub>3</sub> ]  |         | Change in [NH <sup>+</sup> <sub>4</sub> ] |         |      |       |
|---------------|-------------------|---------|------|-------------------------------|---------|---|---------|------|-------|
|               | A                 | В       | С    | A                             | В       | С   | A       | В    | С     |
| ~ 7.5         | n.d.              | n.d.    | n.d. | 0                             | 0       | 0   | -80     | -109 | -89   |
| 6.0           | 467               | 417     | 240  | +40                           | +55     | +36                                       | -61     | -74  | -49   |
| 5.0           | 41                | 29      | 8    | +20                           | +7      | -23                                       | +5      | +1   | -3    |
| 4.5           | 23                | 25      | 0    | -31                           | +40     | +22                                       | -13     | +24  | +27   |
| 4.0           | 15                | 21      | 0    | -29                           | -46     | -62                                       | 0       | -11  | -7    |
| 3.5           | 23                | 14      | n.d. | -36                           | -46     | n.d.                                      | -17     | -2   | n.d.  |
| 3.0           | 18                | 23      | n.d. | -35                           | -36     | n.d.                                      | -18     | -31  | n.d.  |
|               |                   |         |      | NO <sub>3</sub> concentration |         | NH <sup>*</sup> concentration             |         |      |       |
|               |                   |         |      | P                             | L.S.I   | _<br>).                                   | P       | L    | .s.d. |
| DATE          |                   |         |      | 0.059                         | 11      |   | <0.001  |      | 6     |
| STIRRING      | }                 |         |      | 0.862                         | 9       |   | 0.281   |      | 6     |
| PH            |                   |         |      | < 0.001                       | 16      |   | < 0.001 |      | 7     |
| DATE×STIRRING |                   | 0.777   | 16   |                               | 0.591   | 1   | 0       |      |       |
| DATE×PH       |                   | < 0.001 | 28   |                               | < 0.001 | 1   | 3       |      |       |
| STIRRING      | G×PH              |         |      | 0.280                         | 23      |   | 0.293   |      | 3     |
|               | TIRRING           | PH      |      | 0.569                         | 39      |   | 0.140   | 2    |       |

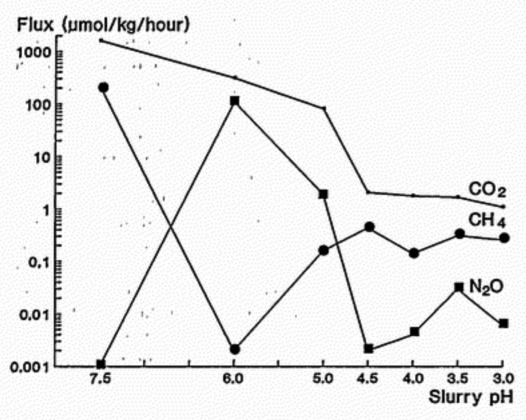


Fig. 5. Fluxes of CO<sub>2</sub>, CH<sub>4</sub> and N<sub>2</sub>O from untreated slurry of pH 7.5 and HNO<sub>3</sub> treated slurries of pH 6.0, 5.0, 4.5, 4.0, 3.5 and 3.0, all from treatment A, on July 7 1991. Note the log scale of the y-axis.

that lowering the pH via the addition of HNO<sub>3</sub> strongly decreased organic C respiration. For CO<sub>2</sub>, the effect was strongest between pH 5.0 and 4.5, whereas even small additions of HNO<sub>3</sub> almost completely inhibited methanogenesis. Flux of N<sub>2</sub>O peaked at pH 6.0. Mean N<sub>2</sub>O fluxes from slurries of pH 6 were about two orders of magnitude higher than those from slurries of pH  $\leq$  5.0 (Oenema et al., 1993). At pH 6.0 the molar C/N ratio of the CO<sub>2</sub> and N<sub>2</sub>O fluxes ranged from 2 to 4, being lowest with stirring treatment A and highest with C. At pH  $\leq$  5.0, the molar C/N ratio was one to two orders of magnitude higher than at pH 6.0.

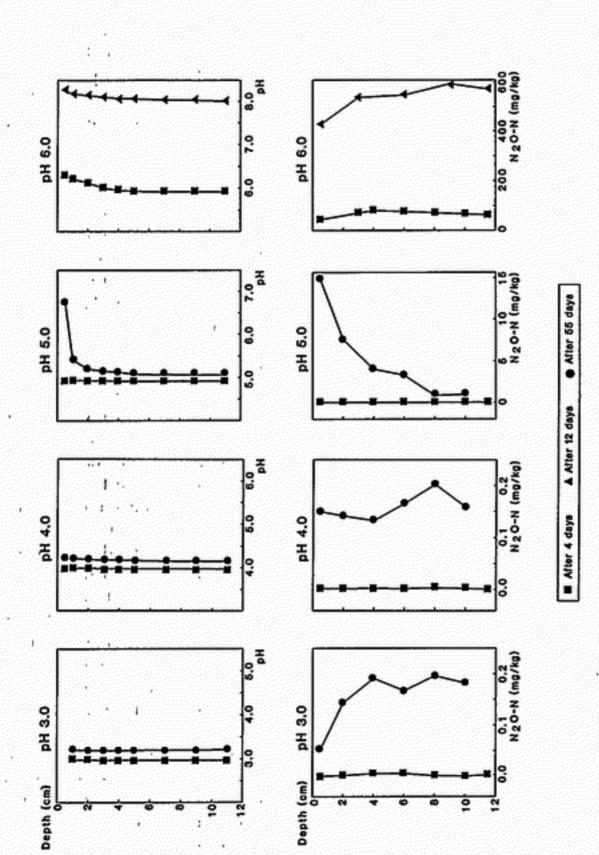
The possible role of nitrification as source of  $N_2O$  was examined in additional batch experiments. Vials containing HNO<sub>3</sub> treated slurry of pH 4.5 and 10 %  $C_2H_2$  in the headspace produced 22.9±0.9 and those without  $C_2H_2$  21.5±1.9  $\mu$ g  $N_2O$ -N kg<sup>-1</sup> d<sup>-1</sup> (n = 5). The similar  $N_2O$  production, in the presence and absence of  $C_2H_2$ , indicates that  $N_2O$  was not produced via nitrification but probably via denitrification.

# Profiles of pH and N2O in slurry

Measurements of pH and  $N_2O$  concentration in unstirred slurries at various depths may provide insight into a possible depth-dependent rate of  $H^+$  consumption and  $N_2O$  production. In slurries of target pH  $\leq$  4.5 the pH remained relatively constant with depth for more than 50 days (Fig. 6). In some treatments with slurry of pH  $\leq$  4.5 small and irregular pH increases were measured in the surface layer (data not shown). The overall pH was increased by about 0.1-0.2 pH units in 55 days. The straight pH profiles suggest that the rate of  $H^+$  consumption was essentially constant with depth or that diffusion could level out differences which may result from a depth-dependent consumption rate. Concentration of  $N_2O$  in slurries of pH 4.5 increased from less than 15  $\mu$ g N kg<sup>-1</sup> after 3 days to about 150  $\mu$ g N kg<sup>-1</sup> after 55 days of incubation.

Significant and rather sudden increases in pH were observed in the surface layer of slurries of pH 5.0 between day 43 (data not shown in Fig. 6) and day 55. This pH increase coincided with an increase in the N<sub>2</sub>O concentration in the surface layer (Fig. 6), suggesting concomitant H<sup>+</sup> consumption and N<sub>2</sub>O production. Because the resolution of our measurements was only about 1 cm, the N<sub>2</sub>O profile in the slurry of pH 5.0 differed from a steady-state diffusion reaction profile of a dissolved constituent typically found in porous media. The high concentrations near the surface and the decreasing concentrations with depth indicate that N<sub>2</sub>O diffused from the surface layers to the atmosphere and to the subsurface. Downward diffusion will continue until the concentration in the subsurface has increased to levels similar to those in the surface layers. Thenceforth, diffusion will be directed to the atmosphere only. This proposed sequence of diffusion, as it follows from the depth and time dependent excursions of pH and N<sub>2</sub>O, also adds to the explanation of the variations in N<sub>2</sub>O flux with time (Oenema et al., 1993).

In slurries of pH 6.0, depth-dependent excursions of pH were measured after 3 and often after only 1 day. After 12 days the pH had increased to about 8.0 irrespective of depth (Fig. 6) and large amounts of N<sub>2</sub>O had accumulated in the slurry. The N<sub>2</sub>O profile of the slurry of pH 6.0 also indicates that a quasi steady state was reached already after 12 days with upward directed diffusion only.



Measurements were carried out 4 and 55 days after homogenization of the slurries by stirring. Note the scale differences of the x-axis of the N2O profiles Fig. 6. Depth profiles of pH (upper half) and N2O (lower half) in HNO3 treated slurries op pH 3.0, 4.0, 5.0 and 6.0 and stirring treatment A. and also that data from slurries of target pH 6.0 are from measurements carried out 4 and 12 days after stirring.

### Discussion

# Cause of the NO3 loss

Basically, there are three possible pathways by which  $NO_3^-$  was removed from the solute in HNO<sub>3</sub> treated slurry: 1) microbiological denitrification; 2) dissimilatory reduction of  $NO_3^-$  to  $NH_4^+$ ; and 3) chemodenitrification. Precipitation of  $NO_3^-$  is not included, because of its high solubility. Immobilization of  $NO_3^-$ , i.e. the assimilatory reduction of  $NO_3^-$  to amines, in biomass is also highly unlikely, because of the low C/N ratio in the slurry (Table 1). Our results provide evidence that microbiological denitrification was a major pathway. The  $NO_3^-$  loss and the  $N_2O$  emission were highest at pH 6.0 and lowest at pH  $\leq$  4.5. The pH-dependance of the  $NO_3^-$  loss is in accordance with the fact that the optimum pH for microbiological denitrification is around 7 and that lowering the pH strongly decreases denitrifying activity (Knowles, 1982).

The HNO<sub>3</sub> treated cattle slurry provides all ingredients needed by denitrifiers: organic C, NO<sub>3</sub> and the (near) absence of O<sub>2</sub> (Tiedje, 1988). The metabolizable organic matter in the slurry serves as energy source and H<sup>+</sup> and electron (e<sup>-</sup>) donor for microbiological denitrification. The overall stoichiometry of the organic matter oxidation via denitrification is given in reactions 1a and 1b, and 2a and 2b of Table 3, with N<sub>2</sub>O and N<sub>2</sub> as the end products, respectively. Such reactions can be used to examine the consistency between H<sup>+</sup> consumption and the NO<sub>3</sub> loss.

Table 3. Stoichiometry of organic matter oxidation in cattle slurry. In the first series of reactions, the molar C:N:P ratio of the organic matter was set at 101:13:1 and the pH of the slurry at about 7. In the second simplified reactions organic matter consisted of CH<sub>2</sub>O only.

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    Nitrate consumption reactions

            (CH<sub>2</sub>O)<sub>101</sub>(NH<sub>3</sub>)<sub>13</sub>H<sub>3</sub>PO<sub>4</sub> + 101HNO<sub>3</sub> = 90CO<sub>2</sub> + 11HCO<sub>3</sub> + 13NH<sub>4</sub> + HPO<sub>4</sub><sup>2</sup> + 50.5N<sub>2</sub>O + 140.5H<sub>2</sub>O
            (CH<sub>2</sub>O)<sub>101</sub>(NH<sub>3</sub>)<sub>13</sub>H<sub>3</sub>PO<sub>4</sub> + 80.8HNO<sub>3</sub> = 90CO<sub>2</sub> + 11HCO<sub>3</sub> + 13NH<sub>4</sub> + HPO<sub>4</sub><sup>2</sup> + 40.4N<sub>2</sub> + 130.4H<sub>2</sub>O
            (CH<sub>2</sub>O)<sub>101</sub>(NH<sub>3</sub>)<sub>13</sub>H<sub>3</sub>PO<sub>4</sub> + 50.5HNO<sub>3</sub> + 11H<sub>2</sub>O = 39.5CO<sub>2</sub> + 61.5HCO<sub>3</sub> + 63.5NH<sub>4</sub> + HPO<sub>4</sub><sup>2</sup>

    Nitrate consumption reaction; simplified reactions

            CH<sub>2</sub>O + HNO<sub>3</sub> = CO<sub>2</sub> + 1.5H<sub>2</sub>O + 0.5N<sub>2</sub>O
            CH<sub>2</sub>O + 0.8HNO<sub>3</sub> = CO<sub>2</sub> + 1.4H<sub>2</sub>O + 0.4N<sub>2</sub>
            CH<sub>2</sub>O + 0,5HNO<sub>3</sub> = 0.5CO<sub>2</sub> + 0.5HCO<sub>3</sub> + 0.5NH<sub>4</sub><sup>4</sup>

    Aerobic oxidition

             CH<sub>2</sub>O + O<sub>2</sub> = CO<sub>2</sub> + H<sub>2</sub>O

    Methanogenesis
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 $CH_2O = 0.5CO_2 + 0.5CH_4$ 

If we assume that the molar C:N:P ratio of the slurry was 101:13:1, as may follow from its composition (Table 1), the overal molar ratio of H<sup>+</sup> consumption and NO<sub>3</sub> loss will vary between 1:0.90 in reaction 1a to 1:0.88 in reaction 1b. The molar ratio of H<sup>+</sup> consumption and NO<sub>3</sub> loss is slightly less than 1:1, because of the mineralization of ammonium and phosphate; the ammonification of 1 mol amine N produces 1 mol alkalinity and the dissociation of 1 mol H<sub>3</sub>PO<sub>4</sub> to HPO<sub>4</sub><sup>2</sup> consumes 2 mol alkalinity. However, a large fraction of total N and P was present in inorganic form (data not shown). This suggests that the molar ratios of C/N and C/P of the decomposing organic matter is higher than 7.8 and 101, respectively, as assumed in reactions 1a and 1b of Table 3. Thus, as another extreme, the stochiometry of the decomposition of N- and P-free organic matter via denitrification is given in the simplified reactions 2a and 2b of Table 3. In these reactions, equivalent amounts of H<sup>+</sup> and NO<sub>3</sub> are consumed, Hence, the molar ratio of H<sup>+</sup> consumption and NO<sub>3</sub> loss will vary between 1:0.88 and 1:1, provided that no other reactions take place than organic matter oxidation via denitrification and mineralization of ammonium and phosphate.

In slurries of pH 6.0 and stirring treatments A, B and C, total NO<sub>3</sub> loss was 427, 362 and 204 mmol kg<sup>-1</sup>, respectively, as follows from the HNO<sub>3</sub> consumption, corrected for the net change in NO<sub>3</sub> concentration (Table 2). The molar ratio of H<sup>+</sup> consumption and NO<sub>3</sub> loss was 1:0.91, 1:0.88 and 1:0.85, respectively. This is close to the overall H<sup>+</sup>:NO<sub>3</sub> ratio of 1:0.90 in reaction 1a and 1:0.88 in reaction 1b of Table 3, assuming that the decomposing organic matter had a molar C:N:P ratio of 101:13:1. However, the changes in NH<sub>4</sub> concentration with time (Table 2) suggest that NH<sub>3</sub> volatilization will also have contributed to H<sup>+</sup> consumption. Volatilization of NH<sub>3</sub> will diminish the total H<sup>+</sup> consumption. The total H<sup>+</sup> consumption, due to organic matter oxidation via denitrification, ammonification and NH<sub>3</sub> volatilization may than be approximated by (all in mmol kg<sup>-1</sup>):

$$H^+$$
 consumption = total  $NO_3^-$  loss +  $\Delta NH_4^+$  (1)

Because H<sup>+</sup> consumption due to mineralization and subsequent dissociation of phosphate is of minor importance, these processes were disregarded in Equation (1). The H<sup>+</sup> consumption unaccounted for (#H<sup>+</sup>) by denitrification, ammonification and NH<sub>3</sub> volatilization may than be approximated by:

$$#H^{+} = \Delta NO_{3}^{-} - \Delta NH_{4}^{+}$$

$$(2)$$

where  $\Delta NH_4^+$  and  $\Delta NO_3^-$  stand for the changes in the  $NH_4^+$  and  $NO_3^-$  concentrations, respectively, between the first and last sampling according to Table 2. In slurries of target pH 6.0, H<sup>+</sup> consumption was larger than accounted for by denitrification, ammonification and  $NH_3$  volatilization (Table 4). In slurries of target pH  $\leq$ 5.0, #H<sup>+</sup> was relatively small and, although mostly negative, may be neglected if we consider the possible variations introduced by all assumptions and analyses involved.

The #H<sup>+</sup> in slurries of target pH 6.0 is puzzling. Total H<sup>+</sup> consumption was more than 25% larger than follows from microbiological denitrification, ammonification and NH<sub>3</sub> volatilization. It suggests that there must have been additional processes

Table 4. Proton consumption unaccounted for according to Equation 2, as a function of pH and stirring treatments A, B and C (mmol kg<sup>-1</sup>). A surplus indicates that more H<sup>+</sup> was consumed than follows from the total NO<sub>3</sub> consumption and the net change in the NH<sub>4</sub> concentration in Table 2.

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|----------------------------------|-------|-----|------|
| pН                               | . · A | В   | С    |
| ~7.5                             | *     |     |      |
| 6,0                              | 102   | 129 | 85   |
| 5.0                              | 15    | 6   | -20  |
| 4.5                              | -18   | -16 | -4   |
| 4.0                              | –29   | -35 | -55  |
| 3,5                              | _19   | -44 | n.d. |
| 3,0                              | -17   | -5  | n.d. |

that contribute to a ratio of H+ consumption/NO3 consumption > 1 and/or processes that have a ratio of H+ production/NO<sub>3</sub> production < 1. Our study was not designed to address these possibilities, but they deserve some discussion. Nitrification does not meet the contraints, because 2 mol H+ is produced for each mol NH4 oxidized to NO3. It is also unlikely that an initial delay in the establishment of an equilibrium, due to the difference in solubility between CO2 and NH3 (Husted et al., 1991) has significantly contributed to #H+, because #H+ was rather small during the first month of the incubation period. Dissolution of previously formed minerals, e.g. carbonates and struvite (Bril & Salomons, 1990) consumes H+ and thus may have contributed to #H+, although the fresh slurry that we used will not have contained large amounts of mineral precipitates yet. Basically, dissimilatory reduction of NO3 to NH4 could also explain that the ratio of H+ consumption and NO3 consumption is larger than 1 (reactions 1c and 2c of Table 3). The dissimilatory reduction of NO3 to NH4 has been shown to occur in highly anoxic and NO3-poor environments, such as digested sludge (Kaspar et al., 1981) and the bovine rumen (Kaspar & Tiedje, 1981). Because NH4 concentrations decreased rather than increased (Table 2), and treated slurries are NO3-rich, dissimilatory reduction of NO3 to NH was probably not an important pathway in these slurries. Chemodenitrification has been put forward in a number of studies to explain unaccountable NO3 losses. Nitrite (NO2), and its undissociated form nitrous acid (HNO2), are key compounds in chemodenitrification, and their chemical decomposition into various volatile N compounds is stimulated generally by acidic conditions (Chalk & Smith, 1983). A possible reaction is: 2 HNO<sub>2</sub> = NO + NO<sub>2</sub> + H<sub>2</sub>O, indicating a 1:1 molar ratio of H<sup>+</sup> consumption and NO<sub>2</sub> consumption. If chemodenitrification occurs concomitant with the oxidation of reduced inorganic compounds (e.g. Sørensen & Thorling, 1991), than the ratio is lower than 1. Relatively high but varying NO2 concentrations were found in slurries of pH 6.0 and low concentrations in slurries of pH ≤ 5.0 (data not shown), which could be explained by the facts that the rate of NO2 formation is related positively to pH and the rate of NO2 decomposition is related negatively to pH (e.g. Van Cleemput & Baert, 1984). This suggests that chemodenitrification cannot be excluded as a possible pathway for NO3 loss from treated slurries. However, we conclude that the large H consumption unaccounted for in slurries of pH 6.0 can not be explained satisfactorily by any of the aforementioned possible processes.

# Denitrification and N2O production

Generally, the ratio of the possible end products of denitrification,  $N_2O/N_2$ , increases with a decrease in pH and with an increase in  $NO_3^-$  concentration (Koskinen & Keeney, 1982). This suggests that more  $N_2O$  in proportion to  $N_2$  will be emitted from slurries of pH  $\leq$  5.0 than from slurries of pH 6.0. If we assume that the total  $NO_3^-$  loss (Table 2) was emitted as  $N_2O$  and  $N_2$ , and combine this with the estimated mean diffusive  $N_2O$  fluxes given by Oenema et al. (1993), it follows that the mean  $N_2O/(N_2O+N_2)$  ratio was 0.5 for slurries of pH 6 and  $\leq$  0.01 for slurries of pH  $\leq$  5.0. This is a surprising result. The relatively low accuracy of the mean  $N_2O$  fluxes and of the  $NO_3^-$  loss for slurries of pH  $\leq$  5.0, because of the relatively low and variable fluxes and  $NO_3^-$  losses, respectively, may have contributed to the apparently reverse effect of pH on the  $N_2O/N_2$  ratio. Emission of other volatile N compounds, such as NO and  $NO_2$ , may also have contributed to the  $NO_3^-$  loss, especially in low-pH slurries. Evidently, the effect of pH on the  $N_2O/N_2$  ratio and on the possible emission of NO and  $NO_2$  from HNO<sub>3</sub> treated cattle slurry needs further study.

As shown in Fig. 5, the molar C/N ratio of the  $CO_2$  and  $N_2O$  emissions was low for slurries of pH 6.0 and much higher for slurries of target pH  $\leq$  5.0. A  $CO_2$ -C/ $N_2O$ -N ratio of about 2 for slurry of pH 6.0 and stirring treatment A would also suggest that the  $N_2O/(N_2+N_2O)$  ratio was about 0.5, if it is assumed that organic carbon was predominantly respired by denitrifiers. An increase in stirring intensity and addition of  $H_2O_2$  (treatment B and C, respectively) increased the  $CO_2/N_2O$  ratio (data not shown), suggesting that aerobic oxidation (reaction 3 of Table 3) became relatively more important, or that the  $N_2O/N_2$  ratio of denitrification decreased. Lowering the pH also increased the  $CO_2/N_2O$  ratio. The apparently positive effect of pH on the  $N_2O/N_2$  ratio and the possible emission of NO and  $NO_2$ , as discussed before, may help to explain that the ratio of  $CO_2/N_2O$  increased as pH decreased. Nonetheless, these findings highlight the need for a better understanding of the effect of pH on the  $N_2O/N_2$  ratio of denitrification in slurry.

Absence of O<sub>2</sub> is generally regarded as a prerequisite for denitrification, even though a number of studies have indicated the persistence of denitrification under the presence of molecular O<sub>2</sub> (Robertson & Kuenen, 1984; Lloyd et al., 1987). In heterogeneous substrates, denitrification seems to be controlled by the rate of O<sub>2</sub> transport to the anaerobic and NO<sub>3</sub> and metabolizable C containing sites of denitrification (Cho, 1982). Surprisingly, in many treated slurries denitrification and N<sub>2</sub>O production were highest at or near the interface of slurry and atmosphere (e.g. Fig. 6). The background of this depth-dependent reaction rate is not yet clear. Apparently, O<sub>2</sub> directly or indirectly stimulated denitrification. It has been suggested that some denitrifiers withstand acidic conditions more easiliy in the presence of small amounts of molecular O2 than in its absence (Oenema et al., 1993). The denitrifying activity in the subsurface slurry may be also partly inhibited by NO<sub>2</sub>, because NO<sub>2</sub> is highly toxic to denitrifiers at relatively low concentrations (Payne, 1981; p. 48-49). Albeit still unclear, the depth-dependant rate of denitrification in HNO<sub>3</sub> treated slurries has important implications for the on-farm treatment of slurry. Regular and complete homogenization of the slurry via stirring appears to be of crucial importance for the control of denitrification. Due to the concomitant consumption of H<sup>+</sup> and NO<sub>3</sub> and its strong pH-dependance, denitrification accelerates, first in the surface layers but subsequently also downwards, when the slurry is not stirred. The accelerating H<sup>+</sup> consumption combined with the relatively low BNC of treated cattle slurry (Fig. 1) may quickly increase the pH to values conducive to NH<sub>3</sub> volatilization, if pH is not quickly adjusted. Regular stirring will level out any gradients in pH and denitrifying activity and will limit the overall NO<sub>3</sub> losses. Local spots with a somewhat elevated pH may also serve for the onset of rapid denitrification. Such 'hot spots' with high N<sub>2</sub>O emissions have also been observed locally in slurry storage basins on dairy farms (unpublished results).

### Conclusions

Treatment of cattle slurry with HNO<sub>3</sub> to lower the pH strongly diminished NH<sub>4</sub><sup>+</sup> losses and CH<sub>4</sub> and CO<sub>2</sub> fluxes during storage. NO<sub>3</sub><sup>-</sup>N losses and flux of N<sub>2</sub>O from these HNO<sub>3</sub> treated slurries had increased compared to untreated slurry. The pH-dependance of the NO<sub>3</sub> loss and the coupling of NO<sub>3</sub> loss, H<sup>+</sup> consumption and N<sub>2</sub>O production (e.g. Table 2 and Fig. 6) indicate that microbiological denitrification was the dominant cause of the NO<sub>3</sub> loss. The H<sup>+</sup> consumption unaccounted for (#H<sup>+</sup>) by denitrification, ammonification and NH<sub>3</sub> volatilization in slurries of pH 6.0 cannot be explained satisfactorily yet with the data now available. This and the depth-dependent denitrification rate in the treated slurries, and the ratio of the flux of N<sub>2</sub>O and N<sub>2</sub> need further study. Lowering of the pH to a value of 4.0-4.5 and regular homogenization of the slurry via stirring are of crucial importance for the succes of the on farm treatment of slurry with HNO<sub>3</sub>.

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